

# Roles/Mechanisms of Adipokines in Development of Metabolic Syndrome

Subjects: [Allergy](#)

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Metabolic syndrome is a cluster of metabolic indicators that increase the risk of diabetes and cardiovascular diseases. Visceral obesity and factors derived from altered adipose tissue, adipokines, play critical roles in the development of metabolic syndrome. Although the adipokines leptin and adiponectin improve insulin sensitivity, others contribute to the development of glucose intolerance, including visfatin, fetuin-A, resistin, and plasminogen activator inhibitor-1 (PAI-1). Leptin and adiponectin increase fatty acid oxidation, prevent foam cell formation, and improve lipid metabolism, while visfatin, fetuin-A, PAI-1, and resistin have pro-atherogenic properties.

adipokine

metabolic syndrome

glucose intolerance

lipid metabolism

## 1. Introduction

Metabolic syndrome is a combination of interrelated conditions that often occur together, including obesity, insulin resistance, glucose intolerance, hypertension, and dyslipidemia <sup>[1]</sup>. Metabolic syndrome is diagnosed as the presence of at least three of the following five characteristics: high waist–hip ratio, high blood pressure, elevated blood sugar level, increased triglycerides (TGs), and low high-density lipoprotein (HDL) cholesterol <sup>[2]</sup>. Metabolic syndrome is important because of its association with an increasing prevalence of diabetes and a higher risk of cardiovascular events such as heart disease and stroke, which have become major public health issues <sup>[3]</sup>. Dysregulation of certain adipokines can promote pathogenic conditions associated with obesity, lipid accumulation, and insulin resistance. These increase the risk of atherosclerosis <sup>[4]</sup>.

## 2. Adipose Tissue as a Critical Endocrine Organ Causing Metabolic Syndrome

Central obesity is the main cause of the etiological cascade of metabolic syndrome. Abnormal fat distribution, rather than adiposity itself, is a more important risk factor for obesity-related disorders <sup>[5][6]</sup>. Recent research has indicated that visceral adipose tissue is 'ectopic fat' that originates from subcutaneous adipose tissue as overflow fat beyond the storage capacity for extra energy <sup>[7][8]</sup>. As 'ectopic fat', visceral adipose tissue is associated with insulin resistance, lipoprotein metabolism, and elevated blood pressure <sup>[9]</sup>. Adipose tissue is an endocrine organ that expresses and secretes various adipokines <sup>[10]</sup>. Adipose tissue includes adipocytes, pre-adipocytes, adipose tissue macrophages, other immune cells, and vascular components. Several factors that are mainly secreted from adipocytes and adipose tissue macrophages contribute to the development of metabolic syndrome <sup>[11][12]</sup>. More

recently, the chronic inflammatory condition that accompanies central obesity has been implicated as a major factor in the development of both metabolic syndrome and its associated pathophysiological consequences [\[13\]](#).

## **| 3. Dysregulation of Adipokines in Metabolic Syndrome**

### **3.1. Leptin**

Leptin is a 16 kDa cytokine primarily produced by white adipose tissue; its secretion increases according to the volume of adipose tissue or adipocytes [\[14\]](#).

### **3.2. Adiponectin**

Adiponectin is a 30 kDa protein that originates from adipose tissue. Adiponectin exists as multimers in plasma and has three major oligomeric forms combined with its collagen domain: a low-molecular-weight trimer, a middle-molecular-weight hexamer, and high-molecular-weight (HMW) 12- to 18-mers [\[15\]](#). HMW adiponectin is a superior biomarker associated with protection against metabolic syndrome as the most potent form in the activation of AMP kinase [\[16\]](#).

### **3.3. Visfatin**

Visfatin is a 52 kDa cytokine that functions like insulin and is expressed in various organs including skeletal muscle, liver, lymphocytes, and adipose tissue [\[17\]\[18\]](#). Visfatin was formerly known as NAMPT (or pre-B-colony-enhancing factor) and is a rate-limiting enzyme that converts nicotinamide to nicotinamide mononucleotide [\[17\]\[18\]](#). Visfatin is also released from visceral adipose tissue, predominantly from macrophages rather than from adipocytes [\[19\]](#).

### **3.4. Fetuin-A**

Fetuin-A, a 64 kDa glycoprotein also known as  $\alpha$ 2-Heremans–Schmid glycoprotein (AHSG), is mainly secreted from the liver and adipose tissue. Fetuin-A contributes to macrophage migration into adipose tissue [\[20\]](#) and augments the expression of proinflammatory cytokines such as IL-6 and TNF- $\alpha$ , while reducing adiponectin expression [\[21\]](#).

### **3.5. Plasminogen Activator Inhibitor-1 (PAI-1)**

PAI-1, a physiological inhibitor of plasminogen activators and vitronectin, is synthesized in adipose tissue.

### **3.6. Omentin-1**

Omentin, also known as Intelectin-1, is a secretory glycoprotein that is highly and selectively expressed in visceral adipose tissue relative to subcutaneous adipose tissue. Both the omentin-1 gene and omentin-2 gene, a homolog

which shares an 83% amino acid identity with omentin-1, are located in the chromosome 1q22-q23 region, which has been previously linked to type 2 diabetes [22].

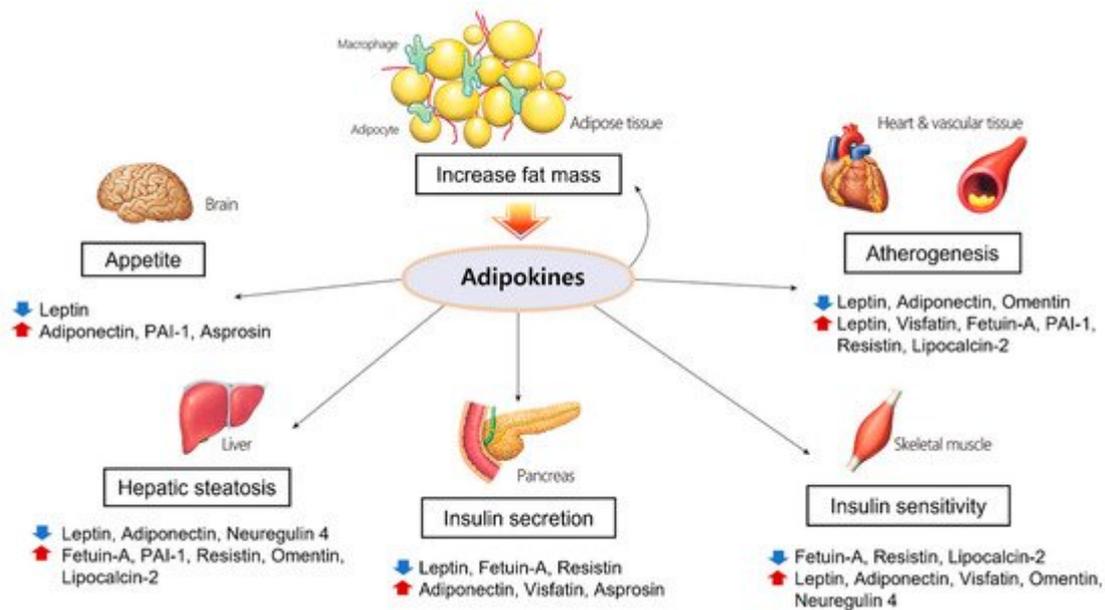
### 3.7. Lipocalin-2

Lipocalin-2 (LCN-2, also called neutrophil gelatinase-associated lipocalin) is a 25 kDa protein that plays a role in the innate immune response to bacterial infection. LCN-2 is expressed in various sites such as liver, kidney, brain, lung, and adipocytes [22].

## 4. Molecular and Cellular Crosstalk in Central Obesity and Metabolic Syndrome

Obesity has generally been considered a risk factor for metabolic and cardiovascular diseases for decades. However, recently, the paradox of obesity has been highlighted from a new perspective in which the location of fat accumulation is the problem rather than the total amount of fat [23]. There are two types of adipose tissue, white and brown, which perform different functions [24]. In humans, white adipose tissue consists mainly of a central intra-abdominal component (visceral adipose tissue) associated with increased metabolic risk, whereas subcutaneous adipose tissue has a protective effect on energy homeostasis and cardiovascular health [25][26]. A previous study showed that human subcutaneous adipose tissue contains larger adipocytes, is less infiltrated by CD68+ and M1-activated cells, and expresses higher levels of cardioprotective adipokines such as adiponectin [27]. Additionally, intraperitoneal implantation of subcutaneous adipose tissue in obese mice prevented glucose intolerance and systemic inflammation [28]. However, the factors that determine visceral or subcutaneous fat distribution remain unknown.

Central obesity induces adipocyte hypertrophy and hyperplasia, macrophage infiltration, endothelial cell activation, and ectopic fat disposition due to excessive energy accumulation. Larger adipocytes are correlated with dysregulated adipokine expression, and hypertrophic adipocytes are prone to producing proinflammatory molecules [29]. In hypertrophic adipose tissue, local hypoxia can occur due to reduced blood flow relative to the size and number of adipocytes, which leads to reduced adiponectin production and increased proinflammatory cytokine expression [30]. Furthermore, obesity not only leads to increased macrophage infiltration in adipose tissue but also triggers their polarization as M1 macrophages producing proinflammatory cytokines and inducible nitric oxide synthase (iNOS) [31]. Through these processes, elevated cytokines and chemokines recruit monocytes that adhere to endothelial cells and elevate the expression of vascular adhesion molecules such as ICAM, VCAM, and E-selectin [32]. This 'vicious cycle' together with chronic inflammation in adipose tissue leads to various complications of metabolic syndrome such as hepatic fibroinflammatory injury, systemic arterial dysfunction, and insulin resistance. In this context, the adipokines and cytokines discussed above play pivotal roles in chronic inflammation, macrophage aggregation, and hypoxia and contribute to the variety of complications called metabolic syndrome (Figure 1).



**Figure 1.** Physiological processes regulated by adipokines in each organ. Adipokines secreted from adipose tissue play important roles in adiposity, glucose and lipid metabolism, and atherosclerosis.

## 5. The Roles and Associated Mechanisms of Adipokines in Cardiovascular Diseases

In a previous study, cardiovascular and all-cause mortality were increased in patients with metabolic syndrome, independent of the baseline cardiovascular disease and diabetes status [33]. Excess weight gain causes an increase in angiotensin II and aldosterone, which regulate renal sodium excretion and play a critical role in blood pressure regulation [34][35]. As an endocrine organ, adipose tissue synthesizes and releases peptides and nonpeptide compounds that have a role in cardiovascular homeostasis. Obesity and enlarged adipose tissue enhance the production of metabolic products that are widely related to atherosclerosis, endothelial dysfunction, hypertension, and dyslipidemia.

### 5.1. Adiponectin

Adiponectin knockout mice exhibited a significantly increased expression of E-selectin, which is implicated in leukocyte rolling and leukocyte adhesion [36]. Aortic ring tissues derived from adiponectin knockout mice showed a decrease in endothelial NOS expression that might cause a defect in vasodilation. This was reversed by treatment with recombinant adiponectin [37]. In another study, adiponectin administration in obese rats increased endothelial NOS by activating the AMPK pathway and promoting NO production. This resulted in the relaxation of the aortic ring [38]. In cultured human umbilical vein endothelial cells, adiponectin showed a protective effect against angiotensin-II-induced vascular endothelial damage [39]. In addition, adiponectin attenuated angiotensin-II-induced NADPH oxidase activation in renal proximal tubular cells [40]. These studies suggest that adiponectin production is closely related to endothelial function in vasodilation.

## 5.2. Leptin

Leptin increased the vasodilatation of rat aortic rings in vitro via a nitric oxide (NO)-dependent mechanism [41], but another study showed that leptin had no effect on hemodynamics, even after blocking NO generation [42]. Moreover, leptin synthesis was found to increase when cultured with angiotensin II adipose cells and rats in vivo [43]. Leptin with adipose-tissue-derived angiotensin II can promote obesity-related hypertension [44]. In an in vitro study using endothelial cells of the human umbilical vein, leptin induced chronic oxidative stress in endothelial cells and promoted atherogenesis [45].

## 5.3. PAI-1

PAI-1, a proinflammatory adipokine, is increased in metabolic syndrome and obesity. PAI-1 has established roles in different pathways for atherosclerosis and cardiovascular risk. An increase in PAI-1 down-regulates tissue plasminogen activator and urokinase activity. This triggers a prothrombotic state and contributes to the development of cardiovascular events. Transient venous thrombosis in the tail and hind limbs was demonstrated in transgenic mice overexpressing PAI-1 [46]. When transgenic mice were generated to express human PAI-1, elevated levels led to spontaneous multiple coronary arterial occlusions and subendocardial infarction in 90% of transgenic mice older than 6 months [47].

## 5.4. Omentin-1

Omentin-1 significantly suppressed foam cell formation in human macrophages. Chronic infusion of omentin-1 into apoE knockout mice reduced atherosclerotic lesions and decreased infiltration of macrophages into atherosclerotic plaques [48]. In a cross-sectional study of type 2 DM patients, omentin-1 was inversely related to arterial stiffness and carotid plaque formation. However, plasma omentin-1 level was significantly elevated in patients with CAD compared to non-CAD patients or healthy volunteers [22]. A recent prospective, 14-year follow-up study of diabetic patients without a previous cardiovascular event observed that higher omentin-1 levels were associated with a higher risk of cardiovascular events, composite incidence of myocardial infarction, stroke, or cardiovascular death, even after adjustment for other risk factors [49].

## 5.5. Lipocalin-2

Cardiac stress induces an increase in LCN-2 expression. Elevated LCN-2 levels were observed in cardiomyocytes of a rat model of post-MI heart failure. High levels of LCN-2 were also detected in the intima of cardiac vasculature after hypoxic stress [50]. LCN-2 expression was increased in atherosclerotic plaques of apolipoprotein E knockout mice compared with control mice [51]. Elevated local levels of lipocalin-2 can mediate cardiovascular function by promoting inflammatory cytokines such as MMP9 and apoptosis [50].

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## References

1. Eckel, R.H.; Grundy, S.M.; Zimmet, P.Z. The metabolic syndrome. *Lancet* 2005, 365, 1415–1428.
2. Alberti, K.G.; Eckel, R.H.; Grundy, S.M.; Zimmet, P.Z.; Cleeman, J.I.; Donato, K.A.; Fruchart, J.C.; James, W.P.; Loria, C.M.; Smith, S.C., Jr.; et al. Harmonizing the metabolic syndrome: A joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. *Circulation* 2009, 120, 1640–1645.
3. Li, X.; Zhai, Y.; Zhao, J.; He, H.; Li, Y.; Liu, Y.; Feng, A.; Li, L.; Huang, T.; Xu, A.; et al. Impact of Metabolic Syndrome and It's Components on Prognosis in Patients With Cardiovascular Diseases: A Meta-Analysis. *Front. Cardiovasc. Med.* 2021, 8, 704145.
4. Kahn, C.R.; Wang, G.; Lee, K.Y. Altered adipose tissue and adipocyte function in the pathogenesis of metabolic syndrome. *J. Clin. Investig.* 2019, 129, 3990–4000.
5. Hu, J.; Yang, S.; Zhang, A.; Yang, P.; Cao, X.; Li, X.; Goswami, R.; Wang, Y.; Luo, T.; Liao, K.; et al. Abdominal Obesity Is More Closely Associated With Diabetic Kidney Disease Than General Obesity. *Diabetes Care* 2016, 39, e179–e180.
6. Fan, H.; Li, X.; Zheng, L.; Chen, X.; Lan, Q.; Wu, H.; Ding, X.; Qian, D.; Shen, Y.; Yu, Z.; et al. Abdominal obesity is strongly associated with Cardiovascular Disease and its Risk Factors in Elderly and very Elderly Community-dwelling Chinese. *Sci. Rep.* 2016, 6, 21521.
7. Smith, U. Abdominal obesity: A marker of ectopic fat accumulation. *J. Clin. Investig.* 2015, 125, 1790–1792.
8. Neeland, I.J.; Ross, R.; Despres, J.P.; Matsuzawa, Y.; Yamashita, S.; Shai, I.; Seidell, J.; Magni, P.; Santos, R.D.; Arsenault, B.; et al. Visceral and ectopic fat, atherosclerosis, and cardiometabolic disease: A position statement. *Lancet Diabetes Endocrinol.* 2019, 7, 715–725.
9. Abate, N.; Garg, A.; Peshock, R.M.; Stray-Gundersen, J.; Adams-Huet, B.; Grundy, S.M. Relationship of generalized and regional adiposity to insulin sensitivity in men with NIDDM. *Diabetes* 1996, 45, 1684–1693.
10. Scheja, L.; Heeren, J. The endocrine function of adipose tissues in health and cardiometabolic disease. *Nat. Rev. Endocrinol.* 2019, 15, 507–524.
11. Thomas, D.; Apovian, C. Macrophage functions in lean and obese adipose tissue. *Metabolism* 2017, 72, 120–143.
12. Appari, M.; Channon, K.M.; McNeill, E. Metabolic Regulation of Adipose Tissue Macrophage Function in Obesity and Diabetes. *Antioxid. Redox Signal.* 2018, 29, 297–312.
13. De Ferranti, S.; Mozaffarian, D. The perfect storm: Obesity, adipocyte dysfunction, and metabolic consequences. *Clin. Chem.* 2008, 54, 945–955.

14. Munzberg, H.; Morrison, C.D. Structure, production and signaling of leptin. *Metabolism* 2015, 64, 13–23.
15. Maeda, K.; Okubo, K.; Shimomura, I.; Funahashi, T.; Matsuzawa, Y.; Matsubara, K. cDNA cloning and expression of a novel adipose specific collagen-like factor, apM1 (AdiPose Most abundant Gene transcript 1). *Biochem. Biophys. Res. Commun.* 1996, 221, 286–289.
16. Hada, Y.; Yamauchi, T.; Waki, H.; Tsuchida, A.; Hara, K.; Yago, H.; Miyazaki, O.; Ebinuma, H.; Kadowaki, T. Selective purification and characterization of adiponectin multimer species from human plasma. *Biochem. Biophys. Res. Commun.* 2007, 356, 487–493.
17. Dahl, T.B.; Holm, S.; Aukrust, P.; Halvorsen, B. Visfatin/NAMPT: A multifaceted molecule with diverse roles in physiology and pathophysiology. *Annu. Rev. Nutr.* 2012, 32, 229–243.
18. Romacho, T.; Sanchez-Ferrer, C.F.; Peiro, C. Visfatin/Nampt: An adipokine with cardiovascular impact. *Mediat. Inflamm.* 2013, 2013, 946427.
19. Curat, C.A.; Wegner, V.; Sengenès, C.; Miranville, A.; Tonus, C.; Busse, R.; Bouloumie, A. Macrophages in human visceral adipose tissue: Increased accumulation in obesity and a source of resistin and visfatin. *Diabetologia* 2006, 49, 744–747.
20. Chatterjee, P.; Seal, S.; Mukherjee, S.; Kundu, R.; Mukherjee, S.; Ray, S.; Mukhopadhyay, S.; Majumdar, S.S.; Bhattacharya, S. Adipocyte fetuin-A contributes to macrophage migration into adipose tissue and polarization of macrophages. *J. Biol. Chem.* 2013, 288, 28324–28330.
21. Dasgupta, S.; Bhattacharya, S.; Biswas, A.; Majumdar, S.S.; Mukhopadhyay, S.; Ray, S.; Bhattacharya, S. NF-kappaB mediates lipid-induced fetuin-A expression in hepatocytes that impairs adipocyte function effecting insulin resistance. *Biochem. J.* 2010, 429, 451–462.
22. Kim, J.A.; Choi, K.M. Newly Discovered Adipokines: Pathophysiological Link between Obesity and Cardiometabolic Disorders. *Front. Physiol.* 2020, 11, 568800.
23. Antonopoulos, A.S.; Tousoulis, D. The molecular mechanisms of obesity paradox. *Cardiovasc. Res.* 2017, 113, 1074–1086.
24. Dhawan, D.; Sharma, S. Abdominal Obesity, Adipokines and Non-communicable Diseases. *J. Steroid Biochem. Mol. Biol.* 2020, 203, 105737.
25. Kwok, K.H.; Lam, K.S.; Xu, A. Heterogeneity of white adipose tissue: Molecular basis and clinical implications. *Exp. Mol. Med.* 2016, 48, e215.
26. Unamuno, X.; Gomez-Ambrosi, J.; Rodriguez, A.; Becerril, S.; Fruhbeck, G.; Catalan, V. Adipokine dysregulation and adipose tissue inflammation in human obesity. *Eur. J. Clin. Investig.* 2018, 48, e12997.
27. Antonopoulos, A.S.; Margaritis, M.; Coutinho, P.; Digby, J.; Patel, R.; Psarros, C.; Ntusi, N.; Karamitsos, T.D.; Lee, R.; De Silva, R.; et al. Reciprocal effects of systemic inflammation and

- brain natriuretic peptide on adiponectin biosynthesis in adipose tissue of patients with ischemic heart disease. *Arterioscler. Thromb. Vasc. Biol.* 2014, 34, 2151–2159.
28. Hocking, S.L.; Stewart, R.L.; Brandon, A.E.; Suryana, E.; Stuart, E.; Baldwin, E.M.; Kolumam, G.A.; Modrusan, Z.; Junutula, J.R.; Gunton, J.E.; et al. Subcutaneous fat transplantation alleviates diet-induced glucose intolerance and inflammation in mice. *Diabetologia* 2015, 58, 1587–1600.
29. Skurk, T.; Alberti-Huber, C.; Herder, C.; Hauner, H. Relationship between adipocyte size and adipokine expression and secretion. *J. Clin. Endocrinol. Metab.* 2007, 92, 1023–1033.
30. Wang, B.; Wood, I.S.; Trayhurn, P. Dysregulation of the expression and secretion of inflammation-related adipokines by hypoxia in human adipocytes. *Pflug. Arch.* 2007, 455, 479–492.
31. Pasarica, M.; Sereda, O.R.; Redman, L.M.; Albarado, D.C.; Hymel, D.T.; Roan, L.E.; Rood, J.C.; Burk, D.H.; Smith, S.R. Reduced adipose tissue oxygenation in human obesity: Evidence for rarefaction, macrophage chemotaxis, and inflammation without an angiogenic response. *Diabetes* 2009, 58, 718–725.
32. Wadey, R.M.; Connolly, K.D.; Mathew, D.; Walters, G.; Rees, D.A.; James, P.E. Inflammatory adipocyte-derived extracellular vesicles promote leukocyte attachment to vascular endothelial cells. *Atherosclerosis* 2019, 283, 19–27.
33. Lakka, H.M.; Laaksonen, D.E.; Lakka, T.A.; Niskanen, L.K.; Kumpusalo, E.; Tuomilehto, J.; Salonen, J.T. The metabolic syndrome and total and cardiovascular disease mortality in middle-aged men. *JAMA* 2002, 288, 2709–2716.
34. Hall, J.E.; do Carmo, J.M.; da Silva, A.A.; Wang, Z.; Hall, M.E. Obesity-induced hypertension: Interaction of neurohumoral and renal mechanisms. *Circ. Res.* 2015, 116, 991–1006.
35. Chandra, A.; Neeland, I.J.; Berry, J.D.; Ayers, C.R.; Rohatgi, A.; Das, S.R.; Khera, A.; McGuire, D.K.; de Lemos, J.A.; Turer, A.T. The relationship of body mass and fat distribution with incident hypertension: Observations from the Dallas Heart Study. *J. Am. Coll. Cardiol.* 2014, 64, 997–1002.
36. Ouedraogo, R.; Gong, Y.; Berzins, B.; Wu, X.; Mahadev, K.; Hough, K.; Chan, L.; Goldstein, B.J.; Scalia, R. Adiponectin deficiency increases leukocyte-endothelium interactions via upregulation of endothelial cell adhesion molecules in vivo. *J. Clin. Investig.* 2007, 117, 1718–1726.
37. Cao, Y.; Tao, L.; Yuan, Y.; Jiao, X.; Lau, W.B.; Wang, Y.; Christopher, T.; Lopez, B.; Chan, L.; Goldstein, B.; et al. Endothelial dysfunction in adiponectin deficiency and its mechanisms involved. *J. Mol. Cell. Cardiol.* 2009, 46, 413–419.
38. Deng, G.; Long, Y.; Yu, Y.R.; Li, M.R. Adiponectin directly improves endothelial dysfunction in obese rats through the AMPK-eNOS Pathway. *Int. J. Obes.* 2010, 34, 165–171.

39. Zhi, Z.; Pengfei, Z.; Xiaoyi, T.; Genshan, M. Adiponectin ameliorates angiotensin II-induced vascular endothelial damage. *Cell Stress Chaperones* 2014, 19, 705–713.
40. Fang, F.; Liu, G.C.; Kim, C.; Yassa, R.; Zhou, J.; Scholey, J.W. Adiponectin attenuates angiotensin II-induced oxidative stress in renal tubular cells through AMPK and cAMP-Epac signal transduction pathways. *Am. J. Physiol. Renal Physiol.* 2013, 304, F1366–F1374.
41. Lembo, G.; Vecchione, C.; Fratta, L.; Marino, G.; Trimarco, V.; d'Amati, G.; Trimarco, B. Leptin induces direct vasodilation through distinct endothelial mechanisms. *Diabetes* 2000, 49, 293–297.
42. Mitchell, J.L.; Morgan, D.A.; Correia, M.L.; Mark, A.L.; Sivitz, W.I.; Haynes, W.G. Does leptin stimulate nitric oxide to oppose the effects of sympathetic activation? *Hypertension* 2001, 38, 1081–1086.
43. Skurk, T.; van Harmelen, V.; Blum, W.F.; Hauner, H. Angiotensin II promotes leptin production in cultured human fat cells by an ERK1/2-dependent pathway. *Obes. Res.* 2005, 13, 969–973.
44. Adamczak, M.; Kokot, F.; Wiecek, A.W. Relationship between plasma renin profile and leptinaemia in patients with essential hypertension. *J. Hum. Hypertens.* 2000, 14, 503–509.
45. Bouloumie, A.; Marumo, T.; Lafontan, M.; Busse, R. Leptin induces oxidative stress in human endothelial cells. *FASEB J.* 1999, 13, 1231–1238.
46. Erickson, L.A.; Fici, G.J.; Lund, J.E.; Boyle, T.P.; Polites, H.G.; Marotti, K.R. Development of venous occlusions in mice transgenic for the plasminogen activator inhibitor-1 gene. *Nature* 1990, 346, 74–76.
47. Eren, M.; Painter, C.A.; Atkinson, J.B.; Declerck, P.J.; Vaughan, D.E. Age-dependent spontaneous coronary arterial thrombosis in transgenic mice that express a stable form of human plasminogen activator inhibitor-1. *Circulation* 2002, 106, 491–496.
48. Watanabe, K.; Watanabe, R.; Konii, H.; Shirai, R.; Sato, K.; Matsuyama, T.A.; Ishibashi-Ueda, H.; Koba, S.; Kobayashi, Y.; Hirano, T.; et al. Counteractive effects of omentin-1 against atherogenesis. *Cardiovasc. Res.* 2016, 110, 118–128.
49. Niersmann, C.; Carstensen-Kirberg, M.; Maalmi, H.; Holleczeck, B.; Roden, M.; Brenner, H.; Herder, C.; Schottker, B. Higher circulating omentin is associated with increased risk of primary cardiovascular events in individuals with diabetes. *Diabetologia* 2020, 63, 410–418.
50. Jang, Y.; Lee, J.H.; Wang, Y.; Sweeney, G. Emerging clinical and experimental evidence for the role of lipocalin-2 in metabolic syndrome. *Clin. Exp. Pharmacol. Physiol.* 2012, 39, 194–199.
51. Hemdahl, A.L.; Gabrielsen, A.; Zhu, C.; Eriksson, P.; Hedin, U.; Kastrup, J.; Thoren, P.; Hansson, G.K. Expression of neutrophil gelatinase-associated lipocalin in atherosclerosis and myocardial infarction. *Arterioscler. Thromb. Vasc. Biol.* 2006, 26, 136–142.

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